

Understanding The Immunological Dynamics of Mycobacterium Tuberculosis Infections

Rani Rai, Education, Glocal School of Education, The Glocal University
Dr. Poonam Lata Middha, (Professor), Glocal School of Education, The Glocal University

ABSTRACT

Humans have been infected with Mycobacterium tuberculosis, the disease's causal agent, for thousands of years. Its capacity to infect, survive, and spread to healthy people is reliant on its ability to elude and exploit host immune responses. The infection cycle frequently leads to an equilibrium state marked by bacterial persistence and immune regulation. According to recent research, different cell types react differently to M. tuberculosis infection, and cellular and intracellular habitats dynamically alter. A variety of lipid and protein effectors that M. tuberculosis possesses can affect inflammatory reactions and macrophage functions. Comprehending the function of distinct bacterial virulence factors in various cellular reservoirs and infection phases is crucial for the creation of innovative treatments, disease indicators, and efficacious vaccinations.

Keywords: Tuberculosis Infections, Immunological Dynamics, Mycobacterium.

1. Introduction

One-fourth of the total populace is contaminated with the respiratory disease Mycobacterium tuberculosis, which has killed a greater number of individuals than some other organism. It comes from environmental mycobacteria, has spent millennia evolving with humans, and has become adept in navigating the human immune system. Comprehending its life cycle is essential for creating therapeutic and preventive vaccinations, innovative treatments, and disease indicators.

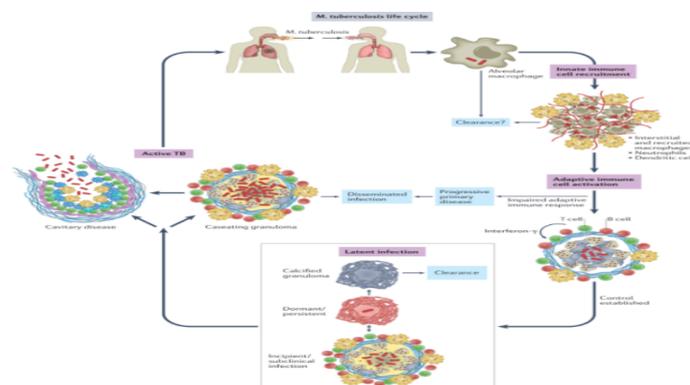


Figure 1: Life cycle of Mycobacterium tuberculosis.

Aerosols from people who have an active lung infection can spread Mycobacterium TB. Dendritic cells, neutrophils, and macrophages are all infected. Thoughts on the innate immune response are uncertain, dendritic cells prime T cells that are specific to antigens. The majority of those infected experience latent illness, whereas 5–10% go on to acquire active TB. The illness presents itself in a variety of ways, eluding immune-mediated removal.

2. Literature Review

Abubakar et al. (2013) conducted a systematic review and meta-analysis to evaluate the duration of protection conferred by bacillus Calmette-Guerin (BCG) vaccination against TB. Their comprehensive analysis synthesized current evidence, providing insights into the effectiveness and longevity of BCG vaccination in preventing TB.

Boom, Schaible, and Achkar (2021) discussed the knowns and unknowns surrounding latent Mycobacterium tuberculosis infection. This review highlights the complexities of latent TB, including factors influencing reactivation and the challenges in diagnosis and treatment. By

addressing gaps in understanding, this review contributes to ongoing efforts to control TB transmission and progression.

Churchyard et al. (2017) provided an overview of what is known about TB transmission, emphasizing the role of various factors such as host susceptibility, environmental conditions, and social determinants. Their review synthesizes current knowledge on TB transmission dynamics, highlighting areas for further research and intervention to reduce transmission rates and TB burden.

Cohen et al. (2018) conducted a study elucidating the role of alveolar macrophages in the early establishment of *M. tuberculosis* infection and dissemination. Their findings demonstrate that alveolar macrophages serve as an initial niche for *M. tuberculosis* replication and contribute to bacterial dissemination within the lung. This study sheds light on early host-pathogen interactions, providing insights into TB pathogenesis and potential targets for therapeutic intervention.

Comas et al. (2013) investigated the evolutionary history of *M. tuberculosis* and its co-expansion with modern humans through an out-of-Africa migration. Their study utilized genetic analyses to trace the origins and spread of *M. tuberculosis* lineages, revealing insights into the long-standing interaction between the pathogen and human populations. This evolutionary perspective enhances our understanding of TB epidemiology and informs strategies for TB control and prevention.

3. *M. Tuberculosis* Establishment of Infection

3.1. The cellular niche of *M. tuberculosis*

Creating a vaccine that prevents infection requires an understanding of the early phases of illness. *M. tuberculosis* is thought to have three bacilli, a low infectious dosage. It comes into contact with alveolar lining fluid, which promotes pathogen uptake and phagocyte death. TB risk is increased by lung surfactant deficiencies brought on either smoking or inflammation. IgM antibodies specific to *M. tuberculosis* may provide protection, according to recent studies.

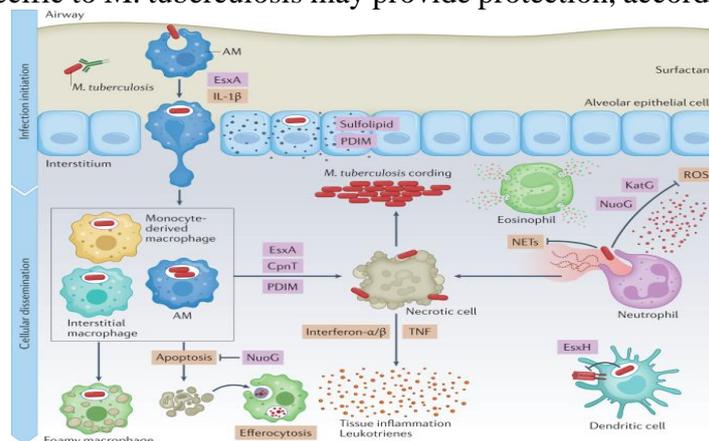


Figure 2: Infection establishment and innate immune evasion by *Mycobacterium tuberculosis*.

Alveolar macrophages (AMs) allow *Mycobacterium tuberculosis* to enter the airways and create a favorable environment for the spread of infection. Depending on the host's IL-1 β creation and the ESX-1 emission framework, tainted AMs move into the lung interstitium. Accordingly, neutrophils produce extracellular neutrophil traps and receptive oxygen species, which just to some extent restrain bacterial development and deteriorate irritation. Certain macrophages have a pro-inflammatory metabolic shift and use antimicrobial mechanisms to kill *M. tuberculosis*, making them more effective at controlling infection. The innate immune system can be subverted by *M. tuberculosis*, and new host-directed therapeutics aimed at

promoting bacterial clearance may be made possible by tactics that tip the infection's scales in favor of restrictive macrophages or increase permissive subsets' antimycobacterial ability.

3.2. Mechanisms of macrophage control

Macrophages identify molecular patterns linked to the *M. tuberculosis* pathogen and initiate antibacterial pathways. These reactions are mediated by cell surface and intracellular pattern-recognition receptors, whereas mycobacterial absorption is encouraged by phagocytic receptors. Pro-inflammatory cytokines and NF- κ B signaling are generated, although host pattern-recognition receptors limit these reactions. After internalizing, macrophages destroy germs by generating antimicrobial peptides, limiting nutrition, and poisoning with heavy metals.

3.3. Mycobacterial immune evasion strategies

For many years, researchers have examined the potential of the bacteria *M. tuberculosis* to weaken macrophage defenses. It inhibits inflammatory signaling, promotes host cell necrosis, and undermines host lysosomal trafficking pathways by the employment of lipid and protein effectors. The lack of significant mitochondrial ROS produced by *M. tuberculosis* infection results in decreased autophagy and NADPH oxidase activity.

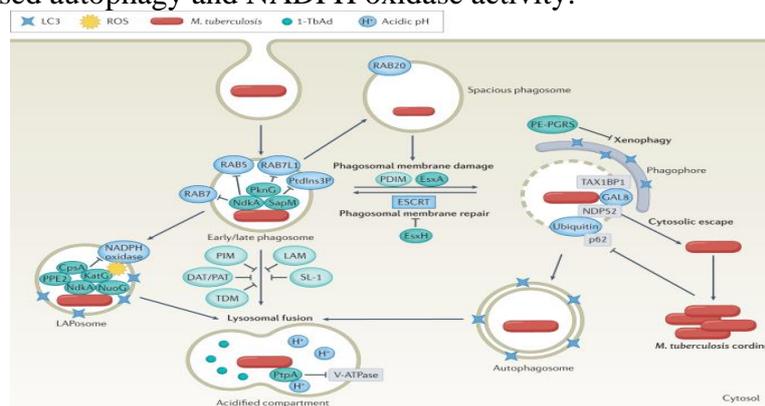


Figure 3: Mycobacterium tuberculosis resides in diverse intracellular compartments.

One membrane-bound phagosome allows Mycobacterium TB to enter the host's body. The host targets this phagosome to aid in the bacterium's removal from the body. The way that virulence factors and host proteins interact determines how infections turn out. By deliberately avoiding phagosomal maturation, Mycobacterium tuberculosis induces oxidative stress and accelerates phagosomal destruction. It can also be directed, in a RAB20-dependent way, toward phagosomes that are roomy. It is possible for *M. tuberculosis* to enter the cytosol and multiply to create cords. Effectors prevent the host from attempting to recapture *M. tuberculosis* that has been exposed to the cytosol in double-membrane autophagosomes. *M. tuberculosis* produces the antacid 1-tuberculosinyladenosine to neutralize pH levels and inhibits vacuolar-type ATPase as defense measures against acidification. Ex vivo investigations have shed light on the ways in which *M. tuberculosis* compromises macrophage functions, but further systems that are physiologically relevant are still required. High-dose therapies (HDTs) targeted at improving mycobacterial clearance can be developed with the aid of an understanding of these immune evasion tactics.

4. Adaptive Immunity Against M. Tuberculosis

Adaptive immune responses are necessary to combat *M. tuberculosis*, as demonstrated by the vulnerability of gene-deleted mice lacking T-cells or MHC class II, as well as lymphopenic HIV patients. Cytokine release and antigen-specific T-cells' direct antibacterial activities are two important aspects of this response. Vaccine development starts with antigen-specific memory T-cells that have a long lifespan. To different antigens, B-cells, $\gamma\delta$ Lymphocytes, and



CD1-confined Immune system microorganisms offer particular responses. Be that as it may, harmful variations to the insusceptible framework are likewise conceivable.

4.1.Kinetics and homing of CD4 T-cells after M. tuberculosis infection

In the mouse disease worldview, CD4 Lymphocyte reactions are fundamental for regulating bacterial multiplication, and their nonappearance can bring about sudden demise. In spite of the fact that CD8 White blood cells are fundamental for vaccination against M. tuberculosis, they can't compensate for a CD4 shortage. One significant trademark that describes the course of M. tuberculosis disease is the beginning of antigen-explicit CD4 Lymphocyte reactions. Notwithstanding the presence of antigen-explicit Lymphocytes, lung antigen-explicit enactment just a brief time after low portion spray disease, demonstrating a postpone in the start of versatile resistant reactions. To restrict intracellular M. tuberculosis replication, CD4 White blood cells draw in with contaminated macrophages. The suitable homing of antigen-explicit CD4 Immune system microorganisms from lymphoid organs to contaminated cells in the lung is vital for the CD4 Lymphocyte reaction to find actual success. For instance, in rhesus macaques, most of antigen-explicit CD4 Immune system microorganisms are situated in the lung parenchyma however are limited to the external lymphocytic sleeve of granulomas, recommending that the distinction between parenchyma-confining and vasculature-limited CD4 White blood cells may not be as critical.

4.2.Quality and specificity of the CD4 T-cell response to M. tuberculosis

For M. tuberculosis contamination, the Lymphocyte reaction is basic, and Th1 and CD8 Immune system microorganism creation of IFN- γ is basic. Qualities incorporate IFNGR1, STAT1, IL12B, and IL12RB are related with hereditary changes that give Mendelian susceptibility to mycobacterial disease (MSMD). Mycobacterial contaminations that return can be brought about by transformations in these qualities. IFN- γ is tracked down in human BAL and reduces after treatment, however it is significant for resistance in creature models. During a disease, versatile resistance might be undermined by an IL-10 deficiency. To appreciate the cycles behind Th1-and IFN-interceded resistance, more examination is required.

Research has shown that FoxP3+ regulatory CD4 Lymphocytes and CD4 White blood cell subsets discharging IL-17 (Th17) support the insusceptible reaction against M. tuberculosis contamination. Th17 cells can work on bacterial control in vivo and intervene assurance without depending on IFN- γ . While FoxP3+ CD4 Immune system microorganisms can limit clear irritation, they can likewise think twice about mycobacterial Lymphocyte reactions and add to ailment. The T-regs' useful job in vaccination against M. Notwithstanding the enactment of versatile resistant reactions, M. tuberculosis delivers a constant contamination in both human and creature models. Bacterial creation of specific antigens during contamination decides antigen-explicit reactions.

4.3.The role of CD8 T-cells in M. tuberculosis infection

Immunity against M. tuberculosis infection depends on CD8 T-cells, which also play a major role in memory responses and reactivation prevention. They release cytokines and effector chemicals, such TNF- α and IFN- γ , that prevent germs from replicating. The HLA-B allele limits the concentration of CD8 T-cell responses on particular epitopes. They participate in antimicrobial activities during active infections and help avoid reactivation during latency.

4.4.Inhibitory receptors during M. tuberculosis infection

Co-inhibitory receptor articulation on Lymphocytes might result from diligent viral diseases, which might affect Immune system microorganism movement. Viral-explicit Lymphocyte reactions that have become practically drained can be resuscitated by obstructing these receptors. Inhibitory receptors like PD-1, CD160, and 2B4 are connected to CD8 White blood cell disappointment during long haul viral diseases. On CD8 White blood cells intended for M. tuberculosis, notwithstanding, their demeanor is negligible. Tim-3 might smother Immune



system microorganism reactions by empowering useful fatigue, despite the fact that PD-1 might be an indication of bacterial burden and CD4 White blood cell initiation. To completely grasp their association in viral resistance, more exploration is required.

4.5. Memory T-cell responses

Following fruitful treatment, memory Lymphocyte reactions have been tracked down in people with LTBI and TB patients. The antigen-particularity, ailment state, and responder recognizable proof all impact these reactions. However, their capacity to give long haul security against *M. tuberculosis* contamination is restricted, memory CD4 and CD8 Lymphocytes add to inoculation against the disease.

4.6. B-cell and antibody responses during *M. tuberculosis* infection

Human granulomas contain B-cells that are engaged with humoral resistance, which supports protection against *M. tuberculosis* disease. Serum from TB patients has been displayed to have antibodies against *M. tuberculosis* proteins, and B-cells can handle irritation to influence how a contamination creates. Furthermore, cytokine discharge by B cells can influence macrophage polarization toward a calming aggregate.

4.7. $\gamma\delta$, CD1-restricted T-cells, and MAIT cells in immunity against *M. tuberculosis*

$\gamma\delta$ Lymphocytes are Immune system microorganisms that recognize non-peptide antigens, for example, phosphoantigens and microbial metabolites and express confined TCR qualities. When presented to monocytes tainted with *M. tuberculosis*, they can increase and respond with mycobacterial heat shock proteins. $\gamma\delta$ Lymphocytes can restrict intracellular replication in macrophages and make up a sizable part of *M. tuberculosis*-receptive Immune system microorganisms in fringe blood. They can also affect DC interaction with T-cells and mediate direct death of *M. tuberculosis*. Glycolipid antigens are delivered by CD1 molecules to T-cells, stimulating T-cell growth and cytokine production. A subgroup of T-cells with innate characteristics that are more prevalent in mucosal tissues are known as mucosal-associated invariant T (MAIT) cells.

5. Conclusion

Understanding how tuberculosis (TB) impairs the functioning of macrophages and T cells, as well as how bacilli take use of the host immune response to create a cellular niche, inflict tissue damage, and facilitate transmission, has advanced significantly over the past ten years of TB study. With many myeloid cell types infected and their virulence tactics emphasized, the intricacy of the cellular response to tuberculosis is showcased. To produce HDTs and successful vaccines, which must overcome myeloid cell malfunction and innate and adaptive immune system failure, it is imperative to comprehend these tactics.

References

1. Abubakar, I., Pimpin, L., Ariti, C., Beynon, R., Mangtani, P., Sterne, J. A., Fine, P. E., Smith, P. G., Lipman, M., Elliman, D., Watson, J. M., Drumright, L. N., Whiting, P. F., Vynnycky, E., & Rodrigues, L. C. (2013). Systematic review and meta-analysis of the current evidence on the duration of protection by bacillus Calmette-Guerin vaccination against tuberculosis. *Health Technology Assessment*, 17, 1–372, v–vi.
2. Boom, W. H., Schaible, U. E., & Achkar, J. M. (2021). The knowns and unknowns of latent *Mycobacterium tuberculosis* infection. *Journal of Clinical Investigation*. <https://doi.org/10.1172/JCI136222>
3. Churchyard, G., Kim, P., Shah, N. S., Rustomjee, R., Gandhi, N., Mathema, B., Dowdy, D., Kasmar, A., & Cardenas, V. (2017). What We Know About Tuberculosis Transmission: An Overview. *The Journal of Infectious Diseases*, 216(Suppl_6), S629–S635.
4. Cohen, S. B., et al. (2018). Alveolar macrophages provide an early *Mycobacterium tuberculosis* niche and initiate dissemination. *Cell Host & Microbe*, 24, 439–446.e4.



5. Comas, I., et al. (2013). Out-of-Africa migration and Neolithic coexpansion of Mycobacterium tuberculosis with modern humans. *Nature Genetics*, 45, 1176–1182.
6. Donald, P. R., et al. (2018). Droplets, dust and guinea pigs: an historical review of tuberculosis transmission research, 1878–1940. *International Journal of Tuberculosis and Lung Disease*, 22, 972–982.
7. Huang, L., Nazarova, E. V., Tan, S., Liu, Y., & Russell, D. G. (2018). Growth of Mycobacterium tuberculosis in vivo segregates with host macrophage metabolism and ontogeny. *The Journal of Experimental Medicine*, 215, 1135–1152.
8. Lee, J., et al. (2020). CD11cHi monocyte-derived macrophages are a major cellular compartment infected by Mycobacterium tuberculosis. *PLoS Pathogens*, 16, e1008621.
9. Mangtani, P., Abubakar, I., Ariti, C., Beynon, R., Pimpin, L., Fine, P. E., Rodrigues, L. C., Smith, P. G., Lipman, M., Whiting, P. F., & Sterne, J. A. (2014). Protection by BCG vaccine against tuberculosis: a systematic review of randomized controlled trials. *Clinical Infectious Diseases*, 58, 470–480
10. Norris, B. A., & Ernst, J. D. (2018). Mononuclear cell dynamics in M. tuberculosis infection provide opportunities for therapeutic intervention. *PLoS Pathogens*, 14, e1007154.
11. Pai, M., et al. (2016). Tuberculosis. *Nature Reviews Disease Primers*, 2, 16076–16076.
12. Pisu, D., et al. (2021). Single cell analysis of M. tuberculosis phenotype and macrophage lineages in the infected lung. *The Journal of Experimental Medicine*. <https://doi.org/10.1084/jem.20210615>
13. Pisu, D., Huang, L., Grenier, J. K., & Russell, D. G. (2020). Dual RNA-Seq of Mtb-infected macrophages in vivo reveals ontologically distinct host-pathogen interactions. *Cell Reports*, 30, 335–350.e4.
14. Roy, A., Eisenhut, M., Harris, R. J., Rodrigues, L. C., Sridhar, S., Habermann, S., Snell, L., Mangtani, P., Adetifa, I., Lalvani, A., & Abubakar, I. (2014). Effect of BCG vaccination against Mycobacterium tuberculosis infection in children: systematic review and meta-analysis. *BMJ*, 349, g4643.
15. Turner, R. D., Chiu, C., Churchyard, G. J., Esmail, H., Lewinsohn, D. M., Gandhi, N. R., & Fennelly, K. P. (2017). Tuberculosis Infectiousness and Host Susceptibility. *The Journal of Infectious Diseases*, 216(Suppl_6), S636–S643.